



# **Technical meeting**

Introduction of Seahorse® technology for studying the metabolism





Seahorse device is equipped with fluorescent probes allowing the measurement of:

- Oxygen consumption rate (OCR) 532/650 nm
- Extracellular acidification rate (ECAR) 470/530 nm

#### A large variety of kits:

- Mito Stress Test: mitochondrial respiration
- Glycolysis Stress Test: glycolysis
- XF Real-Time ATP Rate
- XF palmitate-BSA FAO Subbstrate
- XF Cell Energy Phenotype

Possibility to modulate temperature and measurement times



Agilent Seahorse XFp Analyzer



Agilent Seahorse XFe96 Analyzer



Agilent Seahorse XFe24 Analyzer

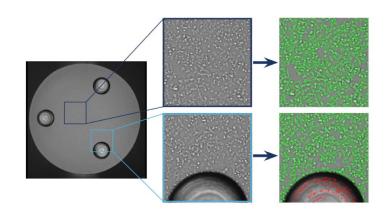
In our laboratory, we have the Seahorse XFe24 analyzer

Sensitivity from 10,000 cells per well

This measurement can be done on different types of sample:

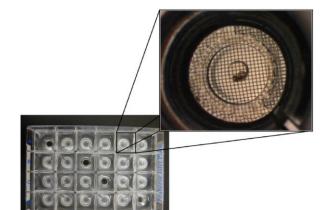
- Cell lines
- Tissues
- Sphéroide
- Zebrafish

Islet plate

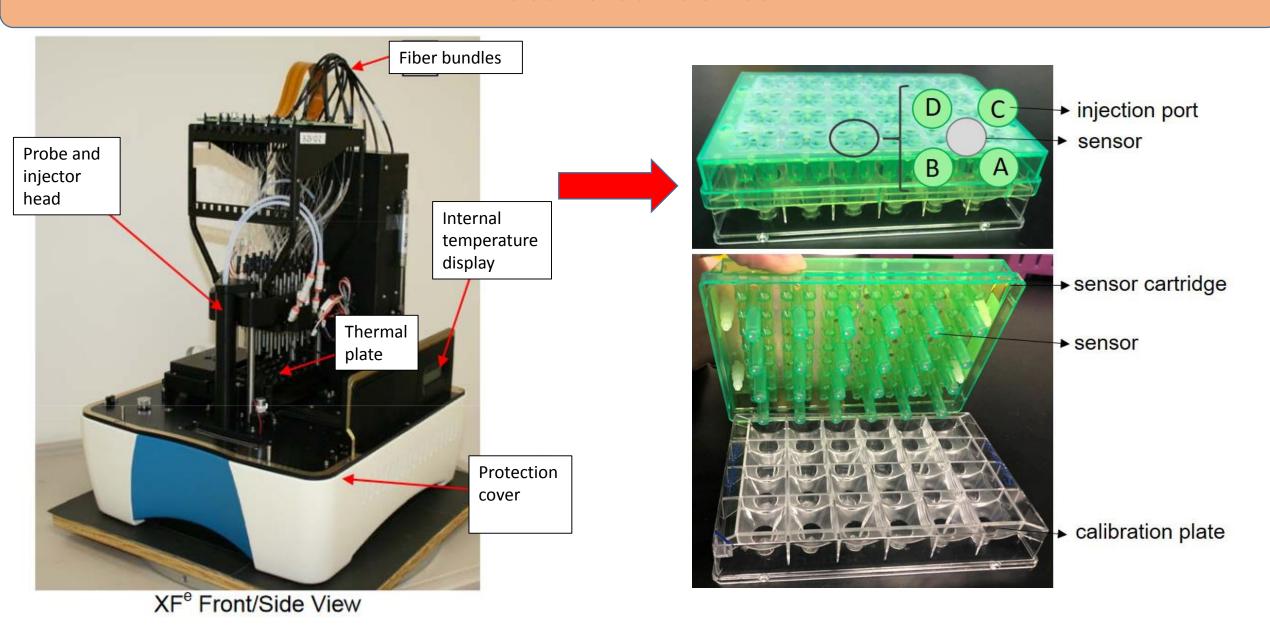


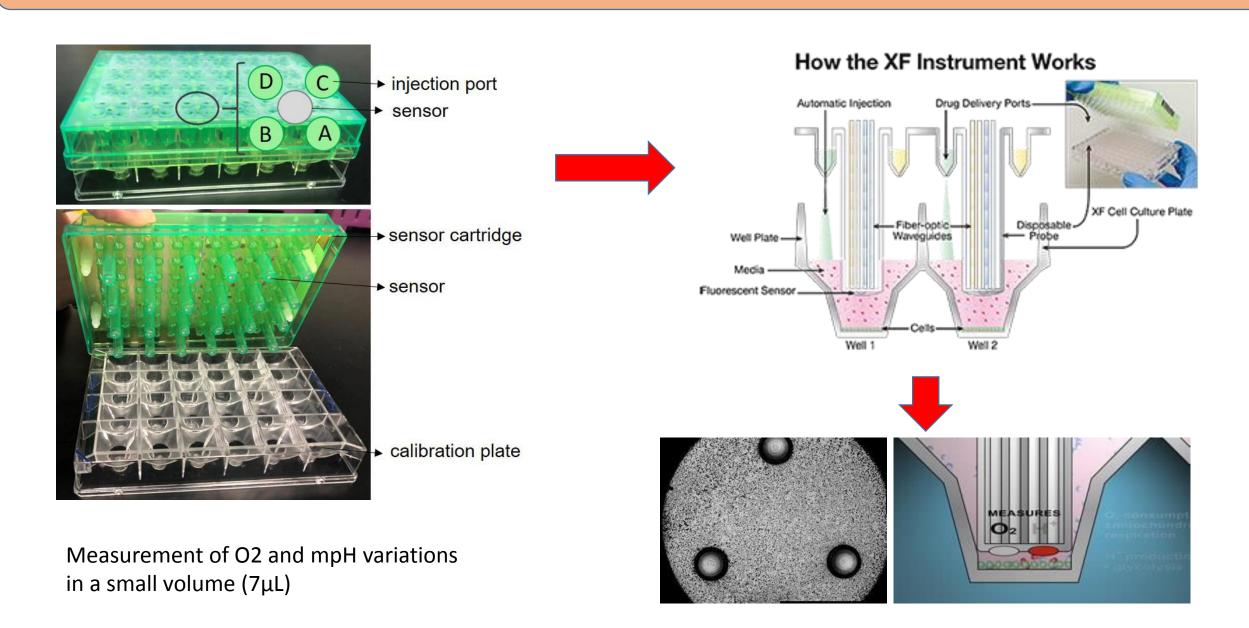
Standard plate

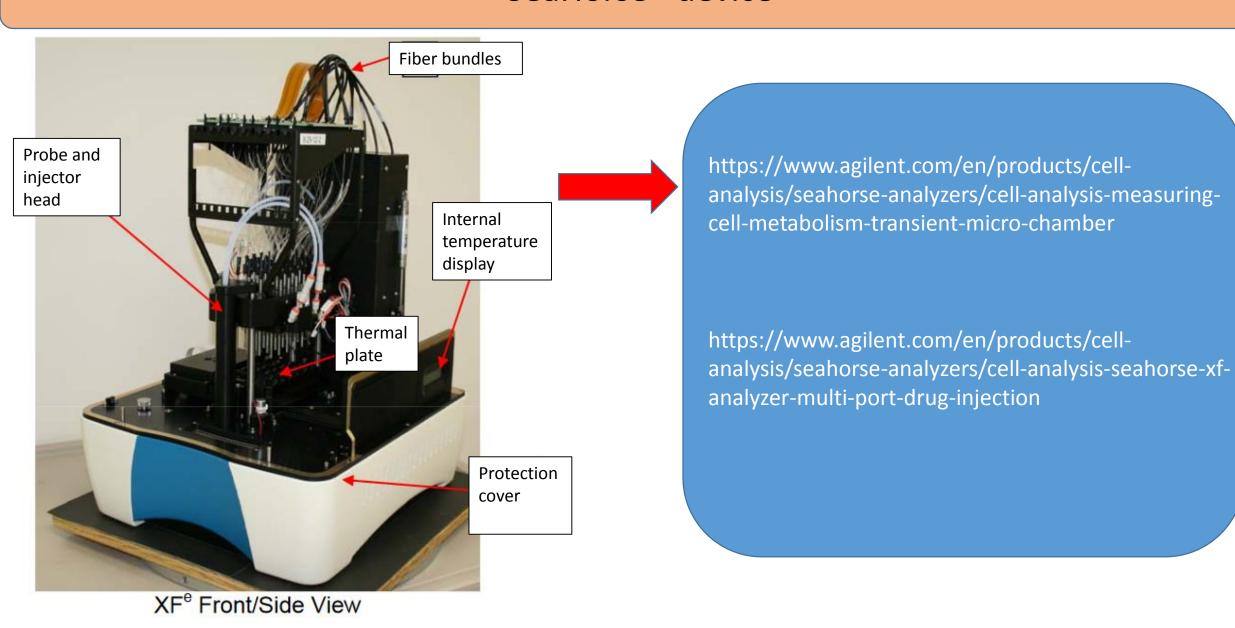


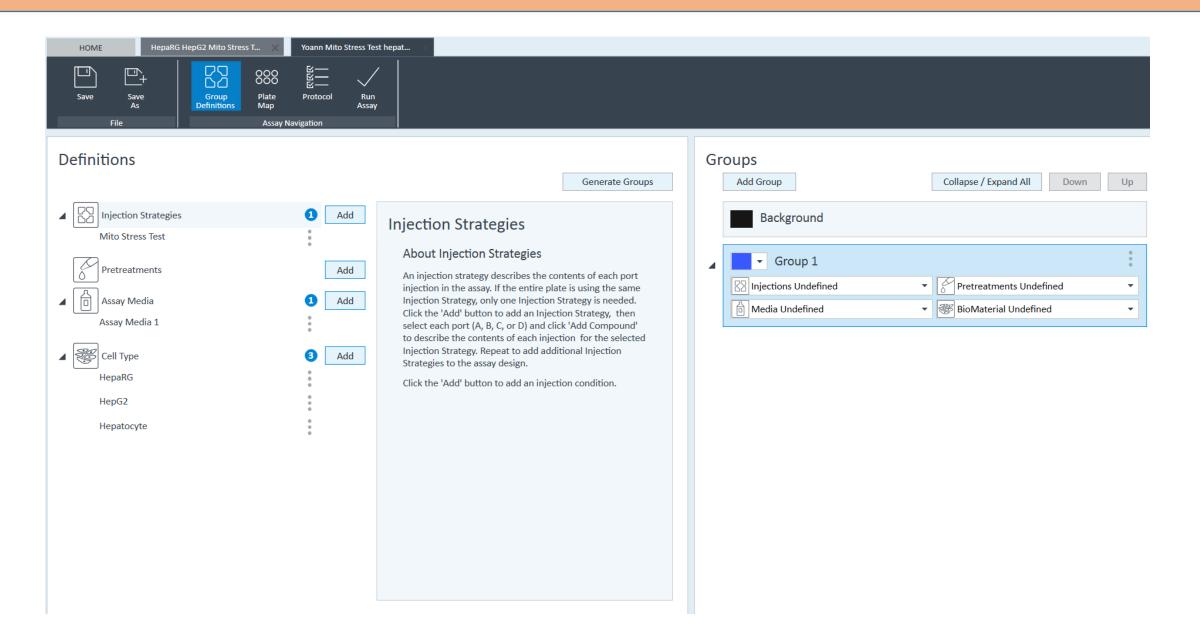


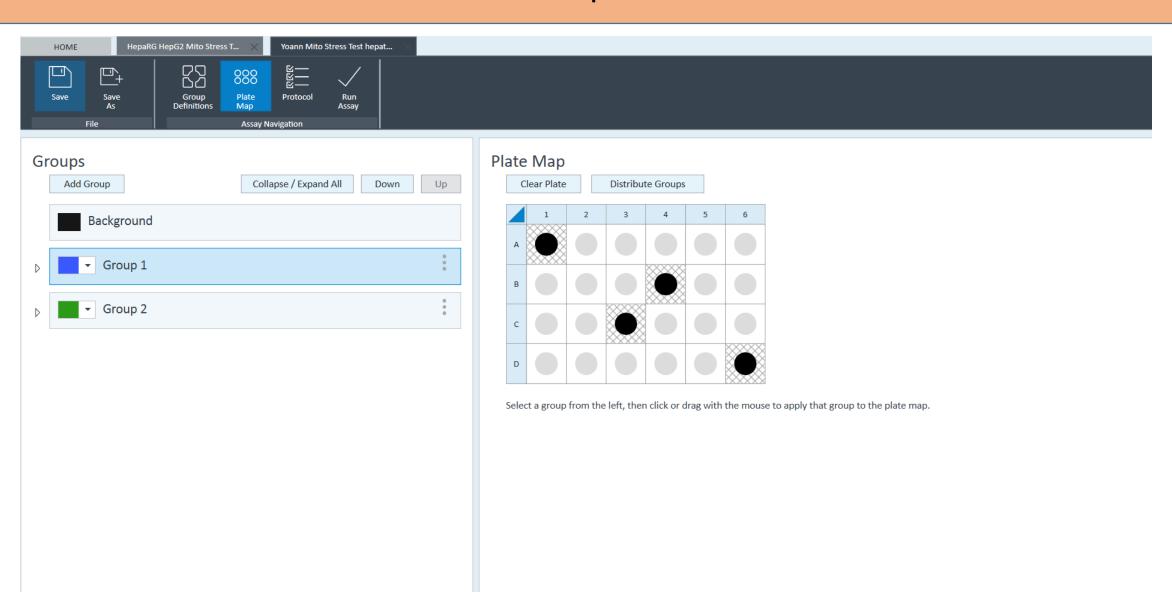
Islet plate

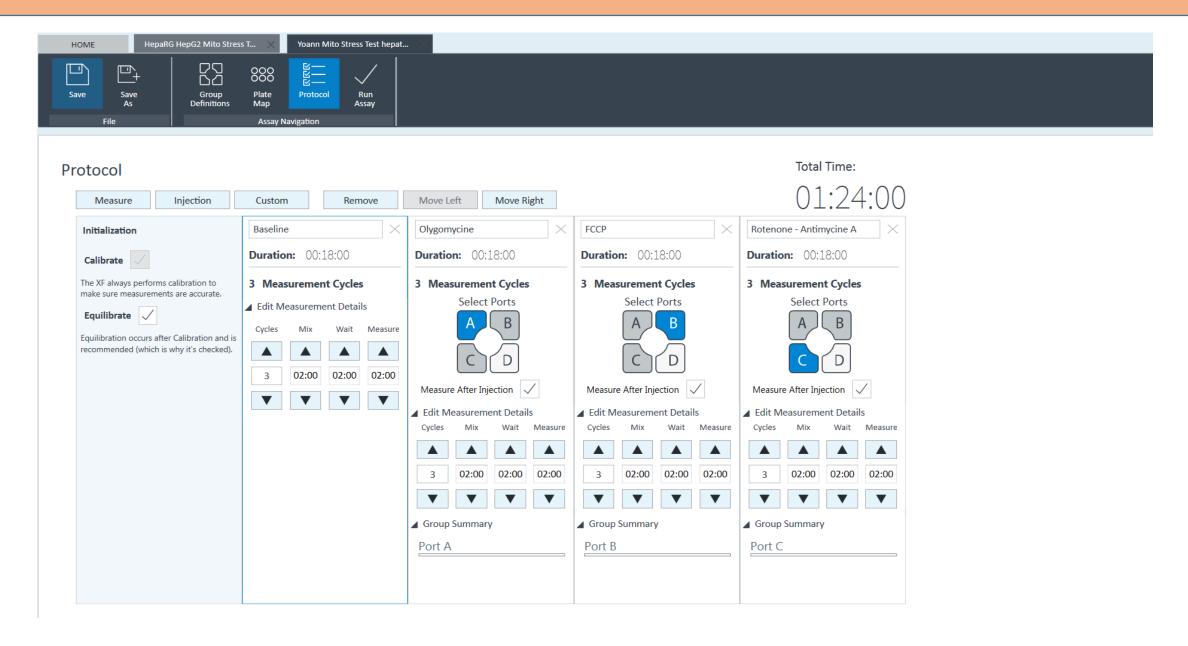


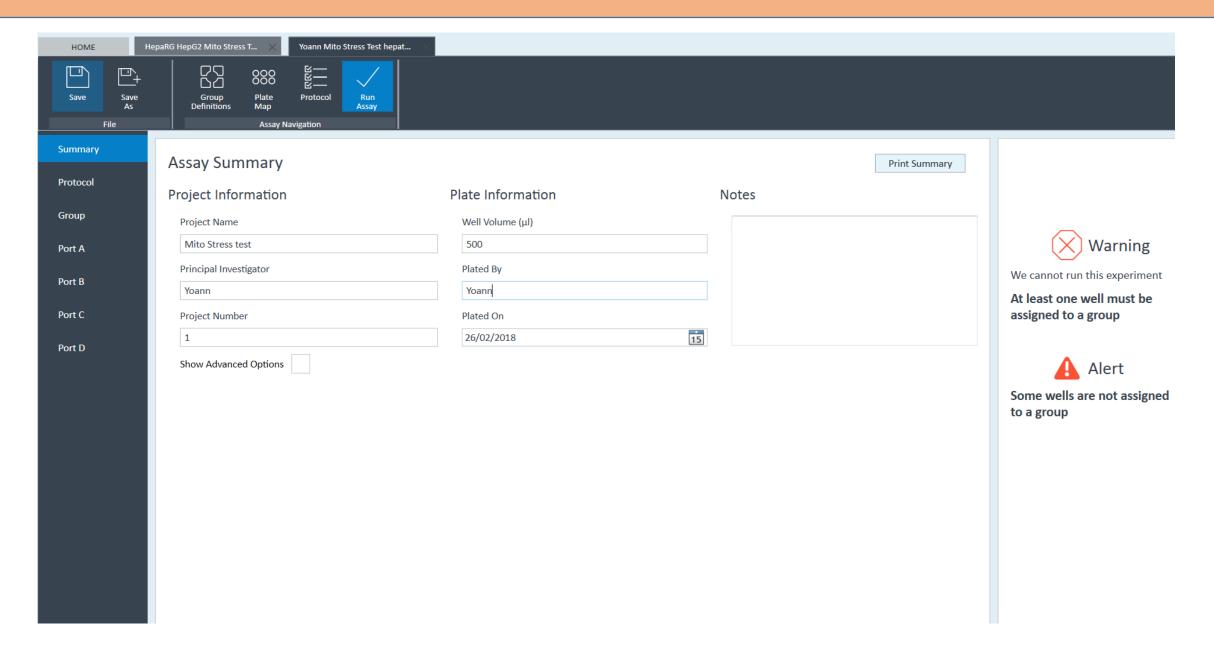




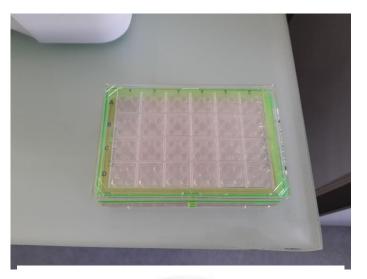








#### Before experiment (Day before)





Hydrate cartridge sensors with XF calibration and place the plate at 37 ° C without CO2 overnight





Prepare the media for the day of the experiment. To do this prepare XF DMEM medium with:

- Glutamine, pyruvate and glucose (Mito Stress Test)
- Glutamine alone (Glycolysis Stress Test)

The pH of the medium must then be adjusted to 7.4

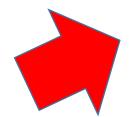
#### Before launch Seahorse device





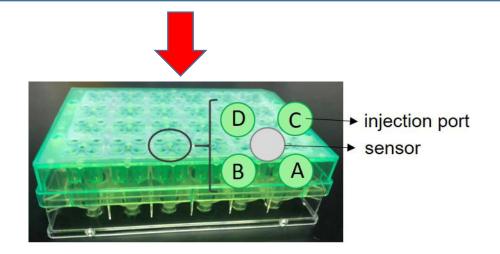
Aspirate the supernatant from the wells and wash with the medium appropriate for the experiment. Then put the plate in the 37 ° C CO2-free incubator for 45 min to 1 hour



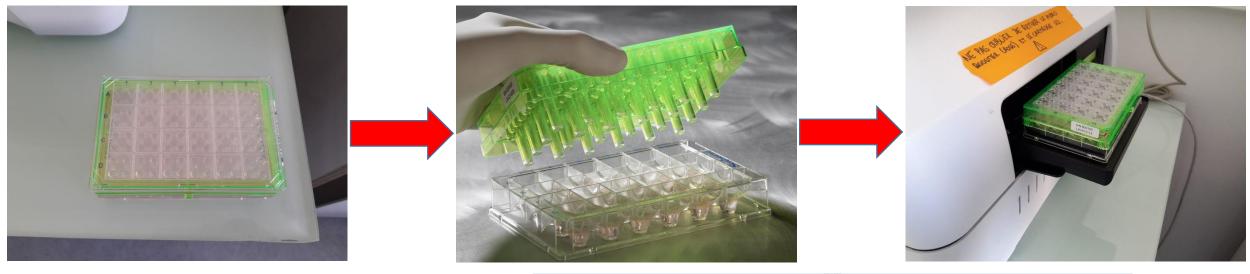


Reconstitute the different molecules with the appropriate medium and load them into the cartridge which was in the incubator at 37 ° C overnight

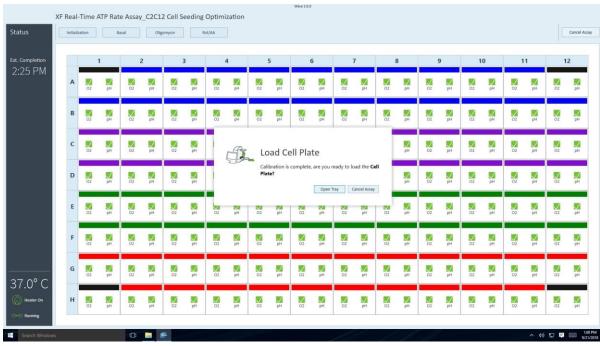




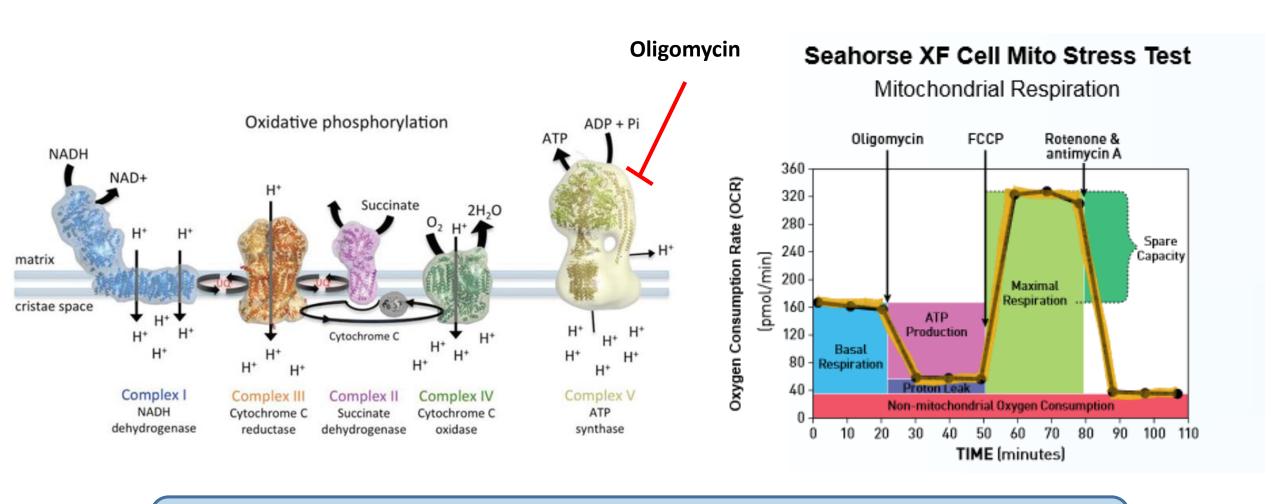
#### Calibration



Remove the cover and the pink plate from the cartridge and put it in the device for pH and O2 calibration

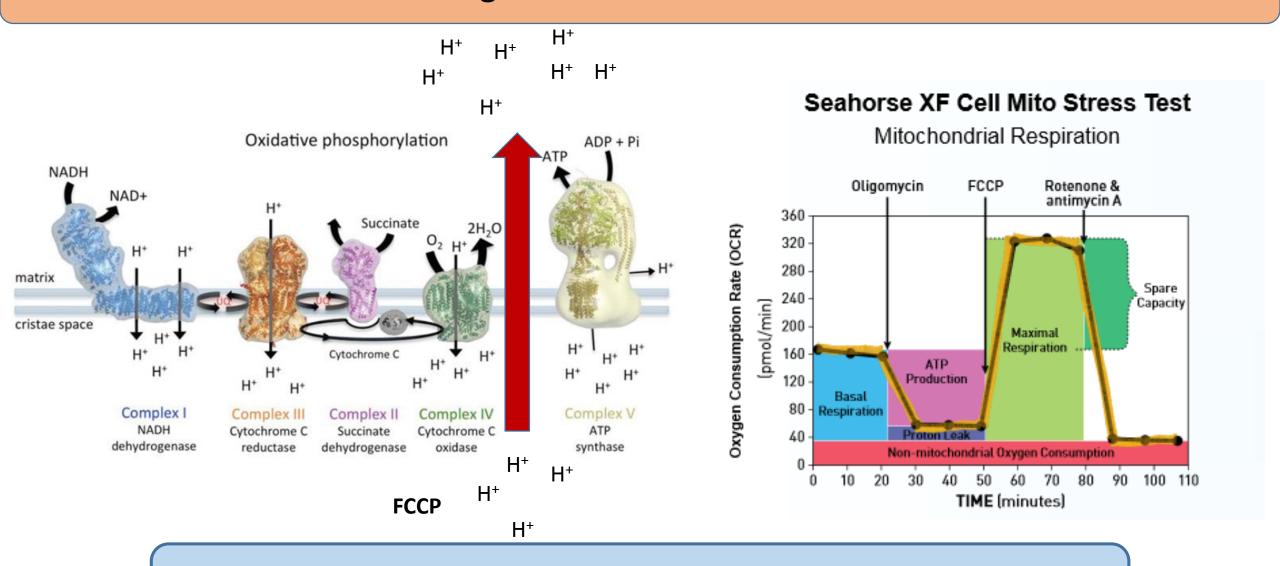


#### During the run: Mito Stress Test



Oligomycin inhibits ATP synthase complex

#### During the run: Mito Stress Test



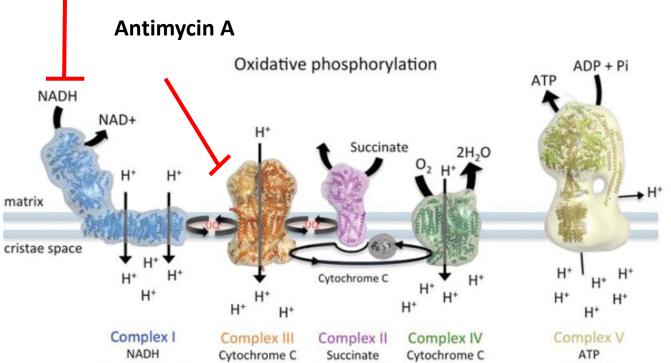
The FCCP will cause a massive entry of protons. The complexes will operate at their maximum capacity

#### During the run: Mito Stress Test

## Rotenone **Antimycin A**

reductase

dehydrogenase

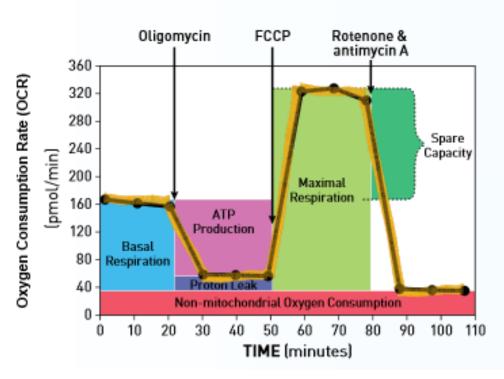


dehydrogenase

oxidase

#### Seahorse XF Cell Mito Stress Test

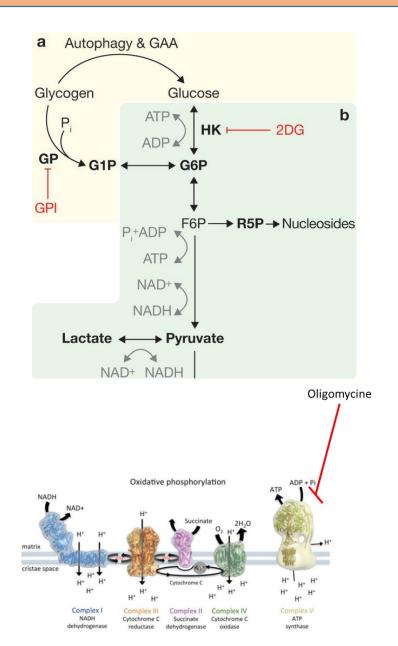
Mitochondrial Respiration

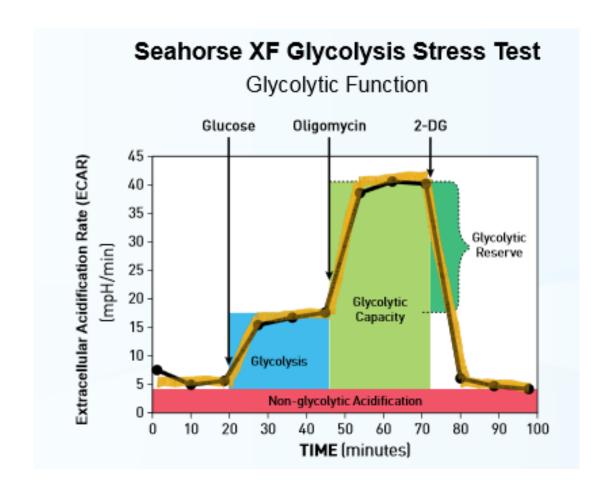


Antimycin A and rotenone inhibit complexes I and III of the respiratory chain causing abrupt arrest of mitochondrial respiration = allows to subtract non-mitochondrial oxygen consumption from all other parameters

synthase

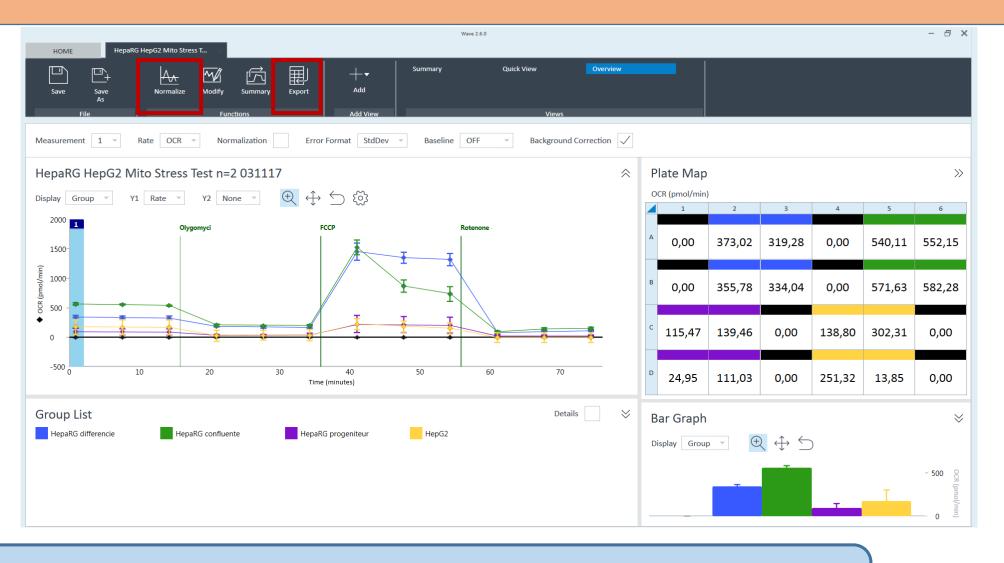
### Other example of kit: Glycolysis Stress Test





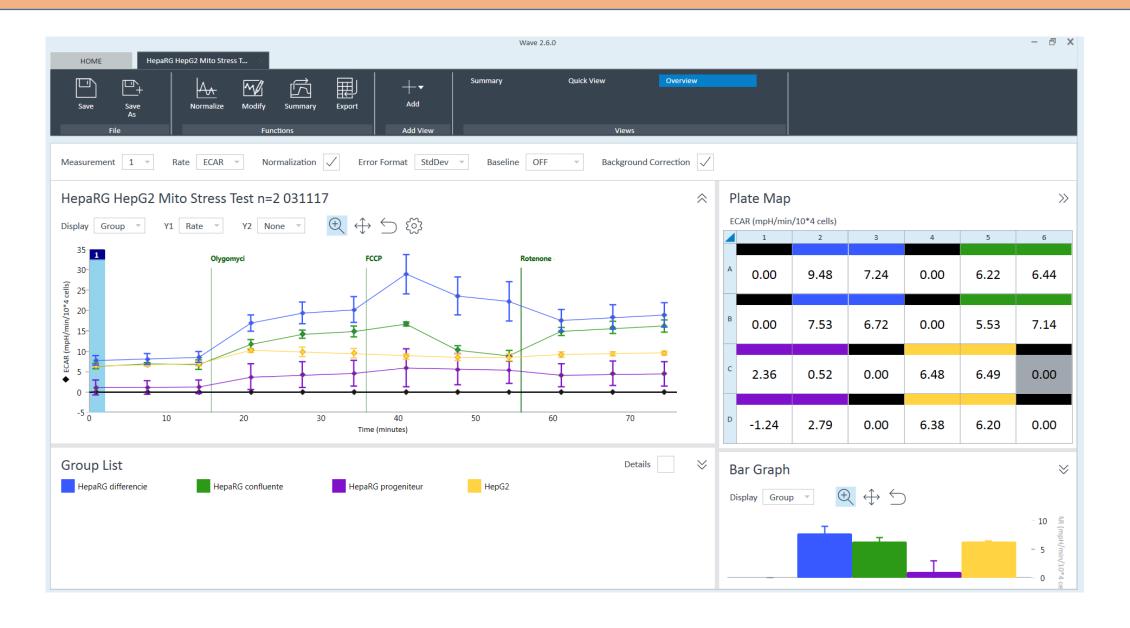
#### Seahorse® Result with Wave software



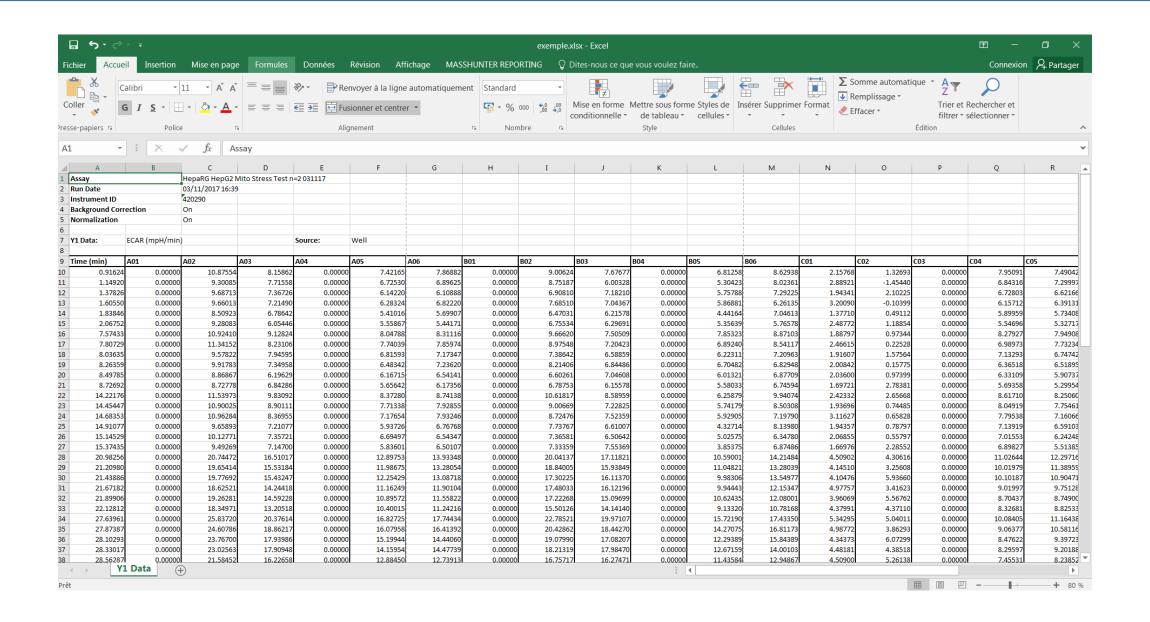


The interface with respiration profiles. We can normalize dataset (protein, cell number...) and export them in an Excel

#### Seahorse® Result



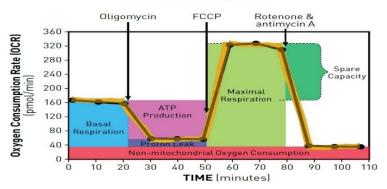
#### Seahorse® Result



#### Seahorse® Result

#### Seahorse XF Cell Mito Stress Test Profile

Mitochondrial Respiration



Parameter Value	Equation
Non-mitochondrial Oxygen Consumption	Minimum rate measurement after Rotenone/antimycin A injection
Basal Respiration	(Last rate measurement before first injection) — (Non-Mitochondrial Respiration Rate)
Maximal Respiration	(Maximum rate measurement after FCCP injection) — (Non-Mitochondrial Respiration)
H+ (Proton) Leak	$(Minimum\ rate\ measurement\ after\ Oligomycin\ injection) - (Non-Mitochondrial\ Respiration$
ATP Production	(Last rate measurement before Oligomycin injection) — (Minimum rate measurement after Oligomycin injection)
Spare Respiratory Capacity	(Maximal Respiration) – (Basal Respiration)
Spare Respiratory Capacity as a %	(Maximal Respiration) / (Basal Respiration) × 100
Acute Response	(Last rate measurement before oligomycin Injection) – (Last rate measurement before acute injection)
Coupling Efficiency	ATP Production Rate) / (Basal Respiration Rate) × 100

	Experimental Group #1						
	Measurement	Assay Well - A1	Assay Well - A2				
	Wicasarcincii	OCR (pmol/min)	OCR (pmol/min)	OCR (pmol/min)	OCR (pmol/min)	Average OCR	
	1	170.02	172.80	169.96	175.99	172.19	
Baseline OCR	2	165.36	163.62	167.00	166.42	165.60	
	3	160.50	158.25	159.44	161.75	159.98	<< Last rate measurement before 1st injection
	4	65.44	53.05	61.53	59.22	59.81	1
Injection 1 - Oligomycin	5	61.80	49.44	59.62	56.94	56.95	
	6	61.93	49.55	59.06	56.04	56.65	<< Minimum rate measurement after Oligo injection
	7	319.58	310.94	312.59	315.98	314.77	
Injection 2 - FCCP	8	327.95	320.32	325.77	330.73	326.19	<< Maximum rate measurement after FCCP Injection
	9	297.17	292.30	301.35	299.68	297.63	
	10	51.77	34.62	44.99	39.45	42.71	
Injection 3 - Rot/AA	11	49.28	33.24	44.13	39.05	41.42	
	12	47.03	31.80	42.66	39.20	40.17	<< Minimum after Rot/AA Injection (Non-Mitochondrial Oxygen Consumpt

 		measurement		4-4:-:
 Last	rate	measurement	ретоге	ist injection

	Individual OCR Values for Assay Wells					
	Measurement	A1	A2	A3	A4	
Baseline OCR	3	160.50	158.25	169.44	161.75	
Injection 1 - Oligomycin	6	61.93	49.55	59.06	56.04	
Injection 2 - FCCP	8	327.95	320.32	325.77	330.73	
Injection 3 - Rot/AA	12	47.03	31.80	42.66	39.20	

Parameter Calculations (per well)		Displayed	Values			
Tarameter Carculations (per wen)	A1	A2	A3	A4	Average	StDev
Non-Mitochondrial Oxygen Consumption	47.03	31.80	42.66	39.20	40.17	6.43
Basal Respiration	113.47	126.45	126.78	122.55	122.31	6.20
Maximum Respiration	280.92	288.52	283.11	291.53	286.02	4.87
H+ (Proton) Leak	14.91	17.75	16.40	16.85	16.48	1.19
ATP Production	98.57	108.70	110.38	105.71	105.84	5.22
Spare Respiratory Capacity	167.45	162.07	156.33	168.98	163.71	5.74

#### Mito Stress Test: interpretation

<u>Basal respiration:</u> Basal oxygen consumption rate (OCR) is the OCR or the rate at which mitochondria function in a cell type under the conditions that you have provided in the culture

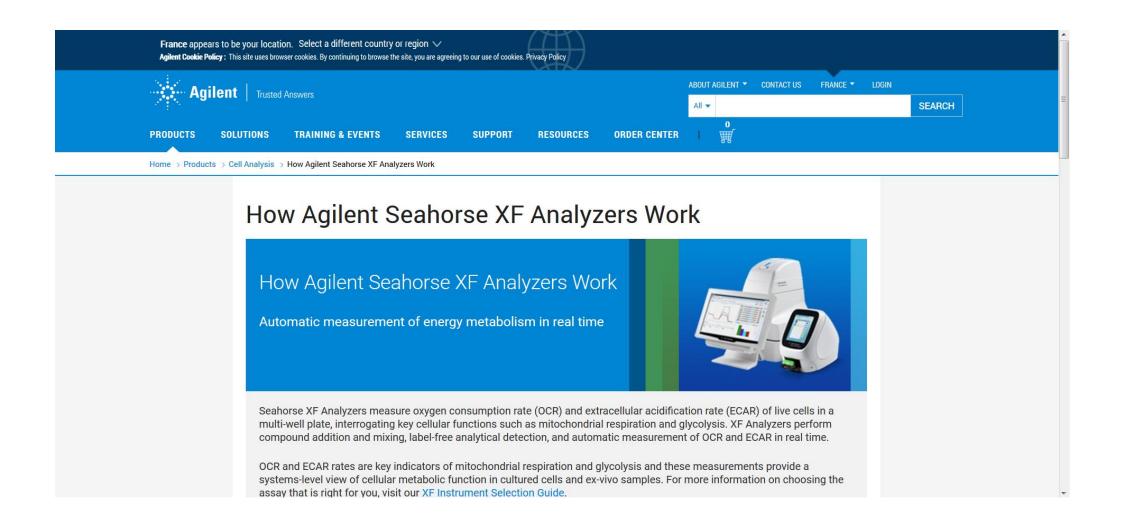
<u>ATP production:</u> Oligomycin sensitive or ATP-dependent OCR is the OCR required to synthesis ATP only at complex V in the mitochondria

<u>Proton leak:</u> This coupling of ATP synthesis and substrate oxidation is not complete, as protons can return to the matrix independently of ATP synthase. The processes by which this occurs are collectively termed "proton leak"

<u>Spare capacity:</u> Spare respiratory capacity or reserve capacity is the difference between maximum OCR after treating the cell with an ionophore and basal OCR, which is an estimate of the potential bioenergetic reserve the cell can call upon in times of stress

Non mitochondrial OCR: This parameter is an index of oxygen consuming processes which are not mitochondrial. In leukocytes, non-mitochondrial OCR is typically attributed to enzymes associated with inflammation, including cyclooxygenases, lipooxygenases and NADPH oxidases, and regarded as negative indicators of bioenergetic health. Non-mitochondrial OCR varies, and typically increases in the presence of stressors, including ROS and RNS and it is well established that mitochondria are a target for the deleterious effects of these reactive intermediates.

#### Agilent website



#### References

Jastroch, Martin, Ajit S. Divakaruni, Shona Mookerjee, Jason R. Treberg, et Martin D. Brand. « Mitochondrial proton and electron leaks ». *Essays in biochemistry* 47 (2010): 53-67. <a href="https://doi.org/10.1042/bse0470053">https://doi.org/10.1042/bse0470053</a>.

Young, Carolyn K. J., et Matthew J. Young. « Comparison of HepaRG cells following growth in proliferative and differentiated culture conditions reveals distinct bioenergetic profiles ». *Cell Cycle* 18, n° 4 (12 février 2019): 476-99. <a href="https://doi.org/10.1080/15384101.2019.1578133">https://doi.org/10.1080/15384101.2019.1578133</a>.

# Thank you for your attention!

